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**SIMULTANEOUS DETERMINATION OF AMOXICILLIN AND RANITIDINE IN
HUMAN PLASMA BY RP-HPLC METHOD**

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ABSTRACT

Gastro duodenal ulcer is a common disease caused by *Helicobacter pylori*. Amoxicillin and ranitidine are usually co prescribed drugs to treat such infections. An RP-HPLC method was developed and validated for the determination of amoxicillin and ranitidine in human plasma simultaneously. For this purpose, chromatographic system comprised of LC-10 AT VP pump and SPD-10 AVP UV/visible detector was used. The separation was best accomplished with mobile phase consisting of 0.02M phosphate buffer: acetonitrile (93:7 v/v, pH 3.0) from Hibar-Purospher star RP-18e (5 µm, 250 x 4.6 mm) column at ambient temperature. The flow rate was 1.0 ml/minute and separation was carried out at 230 nm. The method was found linear over the concentration range of 0.085µg/mL to 20µg/mL for amoxicillin and 0.078µg/ml to 20µg/ml for ranitidine with correlation coefficient of 0.999. The retention times of amoxicillin and ranitidine were about 8 and 11 minutes respectively. The proposed method is accurate, precise, reproducible and economical.

Keywords: Amoxicillin, Ranitidine, RP-HPLC, simultaneous, plasma

1. INTRODUCTION

Helicobacter pylori is the most common cause of gastrointestinal infections worldwide and its eradication significantly reduces the rates of the recurrence and complications [1]. Amoxicillin is a semi synthetic penicillin and commonly prescribed drug in penicillin class due to its good oral absorption. Ranitidine is a H₂ receptor blocker that inhibits production of gastric acid. It is the most commonly used drug to treat peptic ulcer and gastroesophageal reflux disease [2].

For the quantitative determination of amoxicillin trihydrate and ranitidine hydrochloride in plasma, several HPLC methods [3-11], HPLC with diode array detection [12], HPLC/MS [13], HPTLC [14], LC-MS/MS [15,16], UPLC with mass spectrometry methods [17] were presented in the literature. But all these methods were mostly discussed for amoxicillin alone, ranitidine alone or with other combination of drugs.

2. MATERIALS AND METHODS

2.1. Chemicals and reagents

Amoxicillin trihydrate and ranitidine hydrochloride reference standards were gifted by Central Drug Laboratory, Karachi, Pakistan and Indus Pharma (Pvt) Limited respectively. Disodium hydrogen phosphate,

acetonitrile HPLC grade, methanol analytical grade and *ortho* phosphoric acid were purchased from Merck, Darmstadt, Germany.

2.2. Instrumentation and chromatographic conditions

Shimadzu HPLC system equipped with LC-10 AT VP pump and SPD-10 A UV detector loaded with Shimadzu Class GC software was used. Purospher star RP-18e (5 µm, 250 × 4.6 mm) column fitted with a 100 µL loop. Mettler electronic balance and Millipore filtration assembly were also used. pH meter (370 pH meter, Jenway, Europe), analytical balance (Mettler Toledo B204-S, Switzerland), vortex mixer (Whirl Mixer, England), centrifuge (Hereues, Osterode, Germany) swinney membrane filter (Millipore, England), microlitre syringe (Hamilton, Switzerland), and sonicator (LC20H) were used.

2.3. Preparation of mobile phase

The mobile phase having 0.02M phosphate buffer: Acetonitrile (93:7) the pH was adjusted to 3.0 and the flow rate was established at 1.0 ml/minute. The mobile phase was degassed and filtered through 0.45µm filter. The performance was carried out at ambient temperature and detection was made at 230 nm.

2.4. Plasma drug analysis

Blood samples from the human volunteers were collected, plasma was separated and deproteinized by mixing plasma and acetonitrile in a ratio of 1:1, the mixture was vortexed for 5 minutes and centrifuged at 5000 rpm for 8 minutes. Supernatant was then filtered by 0.45 μ membrane filter.

2.5. Method development

The mobile phase which gives best separation was used. The method was optimized by change in composition, pH and flow rate of the mobile phase. Different mobile phases were considered which resulted in broad peaks and more retention time. The finest resolution and retention were achieved when 0.02M phosphate buffer: acetonitrile (93:7) was used and pH was adjusted to 3.0 with *ortho* phosphoric acid.

2.6. Method validation

In the present study, the proposed method was validated by using different parameters like linearity, specificity, accuracy, precision, limit of detection, limit of quantitation, freeze and thaw stability and long term stability of both drugs according to the ICH guidelines.

Specificity differentiates between the active and other substances present in the sample. Specificity was determined to ascertain the separation of amoxicillin and ranitidine with

plasma constituents. It was assessed by injecting the active ingredients and plasma samples.

Linearity of the method was determined by making eleven dilutions of amoxicillin and ranitidine in plasma from 0.039 μ g/mL to 20 μ g/mL. All dilutions were filtered and injected into HPLC separately. The calibration curve was plotted between peak areas versus known concentrations.

Accuracy is the closeness of measured value to the actual value of analytes. It can be found out by repeat sample analysis that have known concentrations of drug. Accuracy can be measured by analyzing at least five aliquots for each concentration.

Precision describes the reproducibility of analytical method. For measurement of precision, five aliquots of each concentration were analyzed. Precision can be divided into intraday and interday. Intraday accuracy and precision is a measure of each accuracy and precision on same day while interday accuracy and precision is a measure of accuracy and precision on different days.

Recovery of an analyte is the response of detector that is obtained from an analyte's concentration when added and extracted from the biologic medium and compared to the response that is found from the actual concentration of the reference standard. It is

the extraction power of the method. The absolute analytical recovery was determined by comparing the peak area of amoxicillin and ranitidine in plasma to that in mobile phase. For both amoxicillin and ranitidine, five aliquots of three different concentrations of 20 µg/mL, 1.25 µg/mL and 0.1562µg/mL were analyzed separately and their percentage recoveries were determined. The relative analytical recovery was determined by comparing the peak area of five aliquots of three different concentrations high, medium and low of the drug (20, 1.25 and 0.1562µg/mL for amoxicillin and ranitidine) with the peak area of actual concentrations. Then percentage recoveries, mean, standard deviations and % CV were calculated.

The freeze and thaw stability of amoxicillin and ranitidine in plasma were determined by preparing fifteen samples each of low (0.085 µg/mL) and high (20µg/mL) concentration. All samples were stored at -20°C. After 24 hours, all samples were thawed; five samples were analyzed and compared with freshly prepared samples. Rests of the samples were re-frozen for next 24 hours. The same procedure was repeated for 2nd and 3rd cycle. Then mean, SD, % CV and percentage accuracy were calculated.

For long term stability, ten samples of the same above mentioned low and high

concentrations of amoxicillin and ranitidine were prepared in plasma and stored at -20°C. To determine the long term stability out of ten, five samples were analyzed at the end of second week and the next five samples were analyzed at the end of fourth week and compared with freshly prepared samples. Statistical parameters i.e. mean, SD, % CV and percentage accuracy were determined.

3. RESULTS AND DISCUSSION

The aim of the present study was to develop and validate an HPLC method for the determination of amoxicillin and ranitidine in human plasma simultaneously.

3.1. Method development

For the determination of amoxicillin and ranitidine simultaneously in human plasma, different mobile phases with different compositions and ratios were tried out like methanol: water (30:70 v/v), phosphate buffer: methanol (90:10 v/v) and phosphate buffer: acetonitrile (93:7 v/v). It was found that methanol containing mobile phases resulted in broad peak with long tail while acetonitrile containing mobile phase give best peak with high resolution.

3.2. Method validation

In the present study, the method was validated according to the ICH guidelines by using different parameters like specificity, linearity, accuracy, precision, limit of

detection, limit of quantitation, freeze and thaw stability and long term stability of both drugs.

3.2.1. Linearity

The standard calibration curve for known concentrations of 20, 10, 5, 2.5, 1.25, 0.625, 0.3125, 0.1562 and 0.085 µg/mL of amoxicillin and 20, 10, 5, 2.5, 1.25, 0.625, 0.3125, 0.1562, 0.085 and 0.078 µg/mL of ranitidine were constructed for linearity, accuracy and precision in plasma and found to be linear with good correlation coefficient ($R^2 = 0.999$) (Table 1, Figure 1 and 2).

3.2.2. Specificity

The chromatograms (Figure 3 and 4) show no interference of amoxicillin and ranitidine with the plasma.

3.2.3. Limit of Detection and Quantification

The lower limit of quantification for amoxicillin and ranitidine were found to be 0.085 µg/mL and 0.078 µg/mL respectively and the limit of detection were 0.078 µg/mL and 0.039 µg/mL for amoxicillin and ranitidine sequentially.

3.2.4. Accuracy

There was no significant difference found between the amount of drug spiked in plasma and the amount recovered and results are presented in table 2.

3.2.5. Precision

Intraday and interday precision was determined by preparing and injecting concentrations of 20, 2.5, 0.3125 and 0.085 µg/mL of amoxicillin and 10, 5, 0.6125 and 0.1562 µg/mL of ranitidine. The mean, SD, precision and accuracy were determined and presented in table 3 and 4. The method was accurate and precise as the results were within acceptable limit.

3.2.6. Robustness

Robustness was analyzed by making deliberate changes in flow rate and pH of mobile phase and the values of %RSD were found less than 2%.

3.2.7. Freeze and thaw stability

Freeze and thaw stability of amoxicillin was determined by subjecting three freeze and thaw cycles to five samples of high concentration (20 µg/mL) and five samples of low concentration (0.085 µg/mL). Similarly for ranitidine, freeze and thaw stability was determined on same number of samples of high concentration (20 µg/mL) and low concentration (0.078 µg/mL). The mean, SD, %CV and % accuracy were calculated (Table 5).

3.2.8. Long term stability

The long term stability was determined on five samples of each high concentration (20 µg/mL) and low concentration (0.085 µg/mL) of amoxicillin and five

samples of each high concentration (20µg/mL) and low concentration (0.078µg/mL) of ranitidine. The mean, SD, %CV and % accuracy were determined for

fresh samples of both drug and then after storage at -20°C at the end of 2nd and 4th week (Table 6).

Table 1: Linearity of amoxicillin and ranitidine in plasma

| Drug | Regression Equation | R ² | LOD (µg/mL) | LOQ (µg/mL) |
|-------------|----------------------|----------------|-------------|-------------|
| Amoxicillin | $y = 15641x - 185.4$ | 0.999 | 0.078 | 0.085 |
| Ranitidine | $y = 42890x + 5964$ | 0.999 | 0.039 | 0.078 |

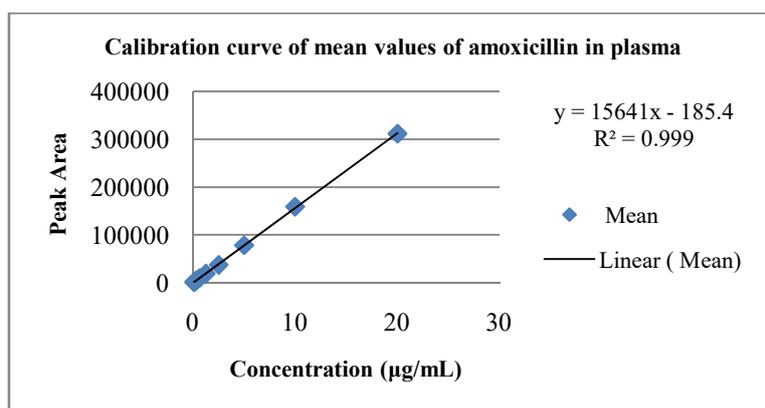


Figure 1: Linearity of amoxicillin in plasma

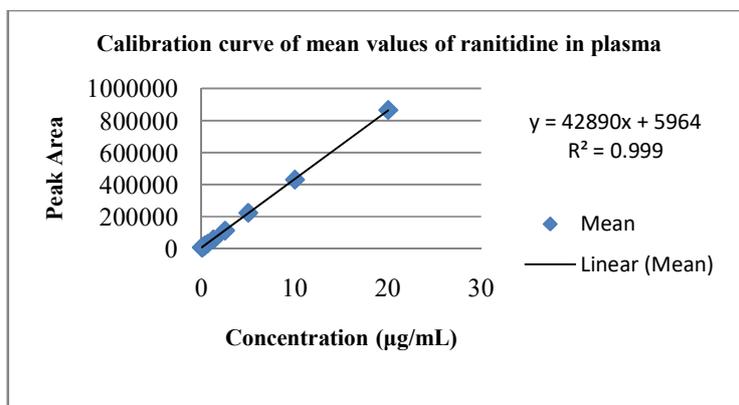


Figure 2: Linearity of ranitidine in plasma

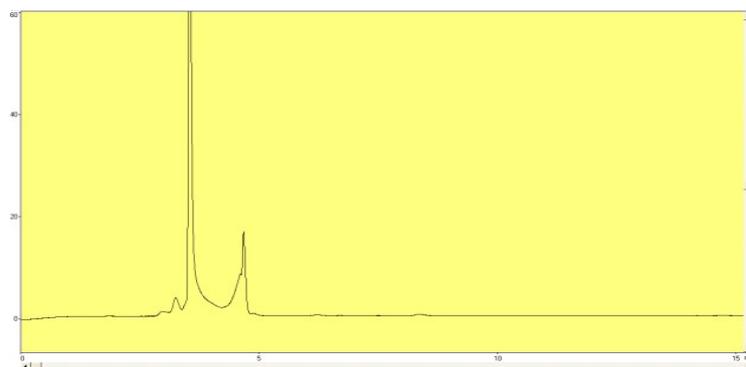


Figure 3: Chromatogram of blank plasma

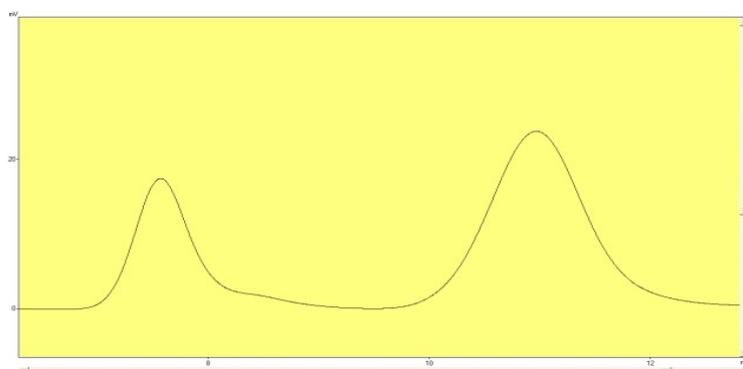


Figure 4: Chromatogram of amoxicillin and ranitidine in plasma

Table 2: Relative analytical recovery of amoxicillin and ranitidine

| RELATIVE ANALYTICAL RECOVERY OF AMOXICILLIN AND RANITIDINE | | | | | | | |
|--|------------|--|------------|-------------------------------------|------------|-------------------------------------|------------|
| | | Actual Concentrations ($\mu\text{g/mL}$) | | | | | |
| | | 20 | | 1.25 | | 0.1562 | |
| | | Measured conc. ($\mu\text{g/mL}$) | % Recovery | Measured conc. ($\mu\text{g/mL}$) | % Recovery | Measured conc. ($\mu\text{g/mL}$) | % Recovery |
| Amoxicillin | Mean (N=5) | 19.669 | 98.343 | 1.2344 | 98.752 | 0.1514 | 96.927 |
| | SD | | 1.350 | | 0.797 | | 1.328 |
| | % CV | | 1.372 | | 0.807 | | 1.370 |
| Ranitidine | Mean (N=5) | 20.023 | 100.119 | 1.245 | 99.664 | 0.1558 | 99.744 |
| | SD | | 0.134 | | 0.537 | | 0.252 |
| | %CV | | 0.133 | | 0.539 | | 0.252 |

Table 3: Intraday accuracy and precision of amoxicillin and ranitidine in plasma
INTRADAY ACCURACY AND PRECISION OF AMOXICILLIN AND RANITIDINE IN PLASMA

| | Theoretical Conc.(µg/mL) | Mean (N=10) | SD | %CV | %Accuracy |
|-------------|--------------------------|-------------|-------|-------|-----------|
| Amoxicillin | 20 | 19.732 | 0.341 | 1.728 | 98.659 |
| | 2.5 | 2.485 | 0.016 | 0.639 | 99.388 |
| | 0.3125 | 0.310 | 0.005 | 1.519 | 99.277 |
| | 0.085 | 0.083 | 0.001 | 1.807 | 97.294 |
| Ranitidine | 10 | 9.843 | 0.120 | 1.217 | 98.433 |
| | 5 | 4.892 | 0.089 | 1.819 | 97.844 |
| | 0.6125 | 0.6084 | 0.004 | 0.630 | 99.331 |
| | 0.1562 | 0.1550 | 0.001 | 0.736 | 99.232 |

Table 4: Interday accuracy and precision of amoxicillin and ranitidine in plasma
INTERDAY ACCURACY AND PRECISION OF AMOXICILLIN AND RANITIDINE IN PLASMA

| Amoxicillin | Day to day | Measured Conc.(µg/mL) | | | |
|-------------|-------------|-----------------------|--------|--------|--------|
| | | 20 | 2.5 | 0.3125 | 0.085 |
| | Mean (N=15) | 19.774 | 2.493 | 0.302 | 0.083 |
| | SD | 0.336 | 0.020 | 0.016 | 0.001 |
| | Precision | 1.698 | 0.799 | 5.185 | 1.006 |
| | % Accuracy | 98.870 | 99.720 | 96.698 | 97.882 |
| Ranitidine | Day to day | Measured Conc.(µg/mL) | | | |
| | | 10 | 5 | 0.6125 | 0.1562 |
| | Mean (N=15) | 9.712 | 4.845 | 0.610 | 0.155 |
| | SD | 0.030 | 0.042 | 0.002 | 0.0003 |
| | Precision | 0.313 | 0.864 | 0.303 | 0.217 |
| | % Accuracy | 97.122 | 96.892 | 99.533 | 99.065 |

Table 5: Freeze and thaw stability of amoxicillin and ranitidine

| FREEZE AND THAW STABILITY OF AMOXICILLIN AND RANITIDINE | | | | | |
|---|-------------------------------|---------------|------------|------------|------------|
| A M O X I C I L L I N | Low concentration 0.085 µg/mL | | | | |
| | | Fresh samples | FT cycle 1 | FT cycle 2 | FT cycle 3 |
| | Mean (N=5) | 0.0848 | 0.08452 | 0.0843 | 0.08382 |
| | SD | 0.0003 | 0.0003 | 0.00029 | 0.00019 |
| | % CV | 0.417 | 0.368 | 0.346 | 0.229 |
| | % Accuracy | 99.765 | 99.435 | 99.176 | 98.612 |
| | High concentration 20 µg/mL | | | | |
| | | Fresh samples | FT cycle 1 | FT cycle 2 | FT cycle 3 |
| | Mean (N=5) | 19.935 | 19.843 | 19.715 | 19.569 |
| | SD | 0.186 | 0.196 | 0.179 | 0.169 |
| % CV | 0.931 | 0.990 | 0.908 | 0.866 | |
| % Accuracy | 99.673 | 99.213 | 98.576 | 97.847 | |
| R A N I T I D I N E | Low Conc. 0.078 µg/mL | | | | |
| | | Fresh samples | FT cycle1 | FT cycle2 | FT cycle3 |
| | Mean (N=5) | 0.0779 | 0.0778 | 0.0774 | 0.0770 |
| | SD | 0.00008 | 0.00009 | 0.0002 | 0.0004 |
| | % CV | 0.107 | 0.115 | 0.268 | 0.592 |
| | % Accuracy | 99.897 | 99.821 | 99.282 | 98.769 |
| | High Conc. 20 µg/mL | | | | |
| | | Fresh samples | FT cycle1 | FT cycle2 | FT cycle3 |
| | Mean (N=5) | 19.925 | 19.891 | 19.721 | 19.576 |
| | SD | 0.096 | 0.100 | 0.150 | 0.232 |
| % CV | 0.481 | 0.501 | 0.761 | 1.186 | |
| % Accuracy | 99.624 | 99.453 | 98.604 | 97.881 | |

Table 6: Long term stability of amoxicillin and ranitidine in plasma
LONG TERM STABILITY OF AMOXICILLIN AND RANITIDINE IN PLASMA

| Sample No. | Low Concentration | | | High Concentration | | | |
|------------------------|-------------------|---------------|---------------|--------------------|---------------|---------------|--------|
| | Fresh Samples | After 2 weeks | After 4 weeks | Fresh Samples | After 2 weeks | After 4 weeks | |
| | | at -20°C | at -20°C | | at -20°C | at -20°C | |
| Amoxicilli n | Mean (N=5) | 0.0848 | 0.0837 | 0.0833 | 19.935 | 19.895 | 19.808 |
| | SD | 0.0004 | 0.0003 | 0.0002 | 0.186 | 0.087 | 0.090 |
| | % CV | 0.417 | 0.309 | 0.230 | 0.931 | 0.436 | 0.453 |
| | % Accuracy | 99.765 | 98.494 | 98.023 | 99.673 | 99.473 | 99.041 |
| Ranitidine | Mean (N=5) | 0.0779 | 0.0775 | 0.0772 | 19.925 | 19.846 | 19.685 |
| | SD | 0.00008 | 0.0002 | 0.0002 | 0.096 | 0.076 | 0.089 |
| | % CV | 0.107 | 0.282 | 0.322 | 0.481 | 0.384 | 0.454 |
| | % Accuracy | 99.897 | 99.410 | 99.076 | 99.624 | 99.232 | 98.424 |

4. CONCLUSION

This RP-HPLC method is suitable for the determination of amoxicillin and ranitidine simultaneously in human plasma. This proposed method is specific, less time consuming and can be used for pharmacokinetic studies of amoxicillin and ranitidine drugs.

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